

October 25th 2021

https://youtu.be/vPLway6jP_I

PRE SYMPOSIUM SEMINAR



"SCHISTOSOMIASIS DIAGNOSTIC RESEARCH AT THE
LUMC: FROM DISCOVERY TO IMPLEMENTATION"

LISETTE VAN LIESHOUT
LEIDEN UNIVERSITY MEDICAL CENTER

"CHALLENGES IN THE DIAGNOSIS OF SCHISTOSOMIASIS:
STILL SEARCHING FOR A GOLD STANDARD"

JOSÉ MAURO PERALTA
FEDERAL UNIVERSITY OF RIO DE JANEIRO



DATE: October 25, 2021 TIME: 14h (Brazilian time)

TRANSMISSION:

FIOCRUZ channel on YouTube.

 www.youtube.com/fundacaooswaldocruz

MODERATOR:

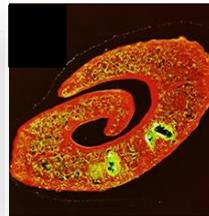
Otávio Sarmiento Pieri



Schistosomiasis diagnostic research at the LUMC: From discovery to implementation



Dr. Lisette van Lieshout
Leiden University Medical Center, Netherlands
Department of Parasitology



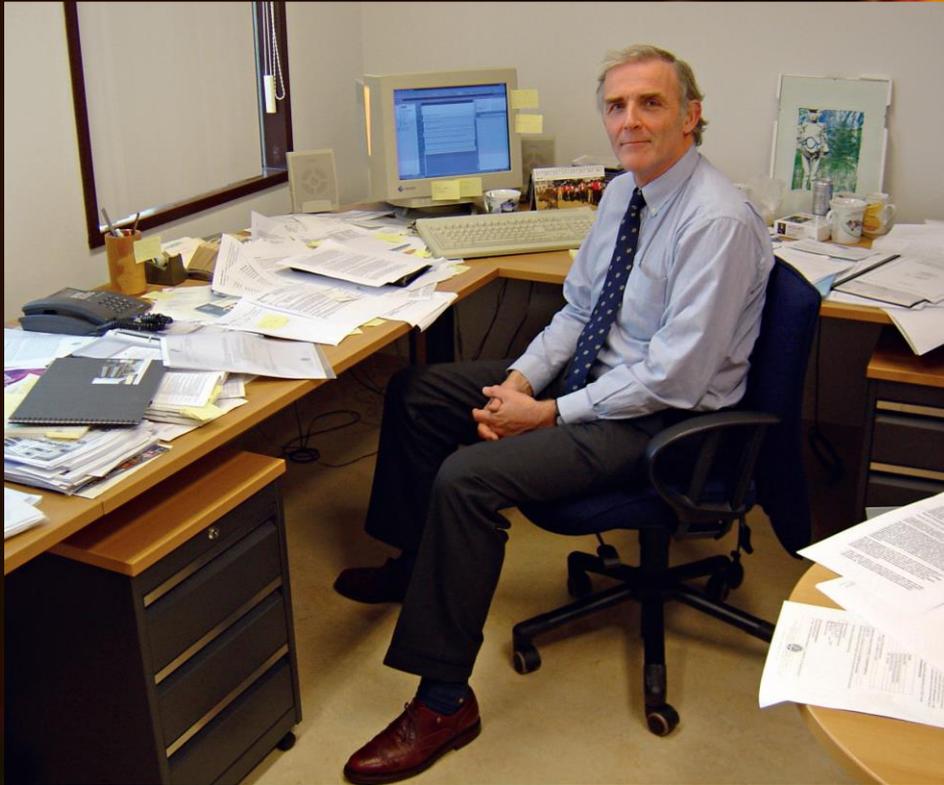
Disclosure:

Department provides the α CAA α CCA McAbs

α CCA to Rapid Medical Diagnostics (South Africa) for urine POC-CCA cassette



In memoriam Professor A.M. Deelder (1947 – 2021)



EXPERIMENTAL PARASITOLOGY 40, 189–197 (1976)

***Schistosoma mansoni*: Demonstration of Two Circulating Antigens in Infected Hamsters**

A. M. DEELDER, H. T. M. KLAPPE, G. J. M. J. VAN DEN AARDWEG,
AND E. H. E. M. VAN MEERBEKE

*Laboratory of Parasitology, University of Leiden,
Rapenburg 33, Leiden, The Netherlands*

(Accepted for publication 27 October 1975)



Laboratory diagnosis of schistosomiasis

Microscopy

- Endemic regions: urine /10 mL; stool Kato-Katz
- WHO reference standard, quantify
- High specificity, but lacks sensitivity
- Simple, but operator dependent



Lack of sensitivity -> repeated

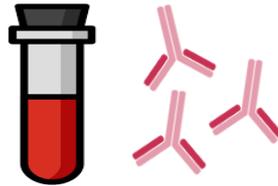


Non-microscopy laboratory diagnosis of schistosomiasis – LUMC research

1. DNA detecting tests (NAAT)



2. Serology (Ab testing)



3. Circulating antigens

- Commercial POC-CCA
- In-house UCP-LF CAA

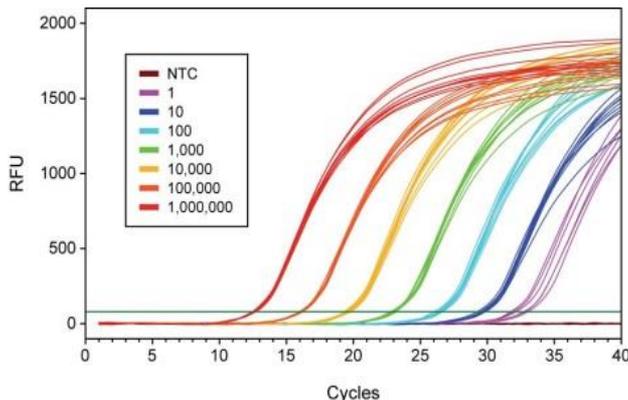


DNA detection: NAAT-based diagnosis

- LUMC: DNA in excreta (faeces, urine, sperm, swaps, lavage)
- (Others: cfDNA: blood-based PCR => urine-excretion)
- Real-time PCR, semi-quantification options (Ct-value)
- Highly specific by design
- Highly sensitive (but not always 100%) – proxy for egg excretion
- Rapidly cleared (<weeks) after treatment



(-) High-tech procedure



*LUMC: ITS-2, genus (S.m./S.h.) Obeng et al., 2008
Routine diagnostics since 2007 (ISO 15189)*

*Participation in proficiency testing (EQAS)
Cools et al. (2020) PLoSNTD*



Female Genital Schistosomiasis

Need for user-friendly diagnosis

Zambia

2.5% Vaginal Swab = 15/603

3.3% Cervical Swab = 20/603

4.0% Any Swab = 24/603

2.7% Cervicovaginal Lavage (CVL) = 14/527

70% of those with FGS had active *Schistosoma* infection* (21/30)

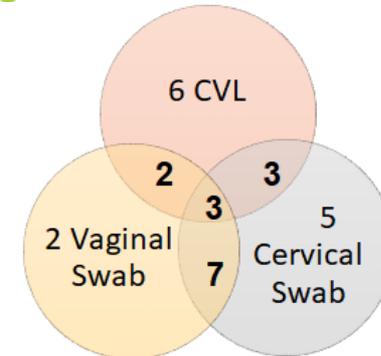
*defined as positive **urine** PCR, CAA, or microscopy

RESEARCH ARTICLE

PLOS NEGLECTED TROPICAL DISEASES

Genital self-sampling compared with cervicovaginal lavage for the diagnosis of female genital schistosomiasis in Zambian women: The BILHIV study

Amy S. Sturt^{1*}, Emily L. Webb², Comfort R. Phiri³, Tobias Mweene³, Namakau Chola³, Govert J. van Dam⁴, Paul L. A. M. Corstjens⁵, Els Wessels⁶, J. Russell Stothard⁷, Richard Hayes², Helen Ayles^{1,3}, Isaiah Hansingo⁸, Lisette van Lieshout⁴, Amaya L. Bustinduy¹

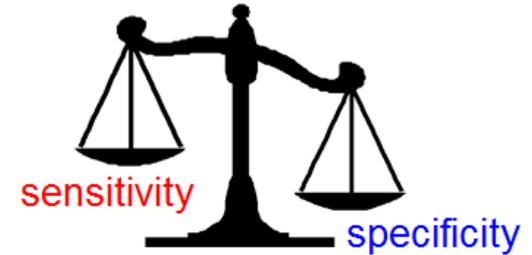
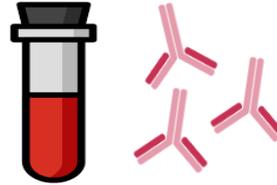


Serology: anti-*Schistosoma* antibody detection

Commercial tests/in-house tests

Specificity??

Sensitivity??



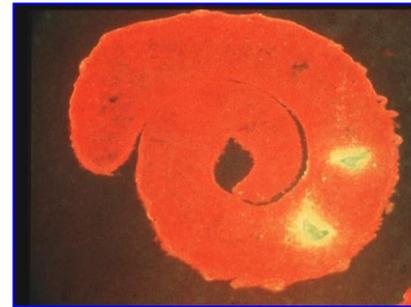
No correlation with intensity of infection/clinical symptoms

Not for evaluation of treatment

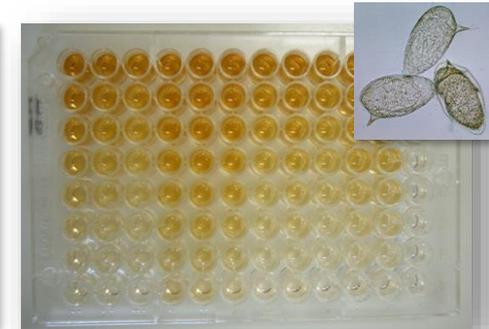
LUMC: in-house tests; ISO 15189

IFA – IgM α -AWA [Nash et al., 1978 AJTMH](#)

ELISA – IgG α -eggs (SEA)



IgM - IFA:
Rossman's fixed male
S.m. adult worms



IgG - ELISA:
crude *S.m.* soluble egg
antigen (SEA)

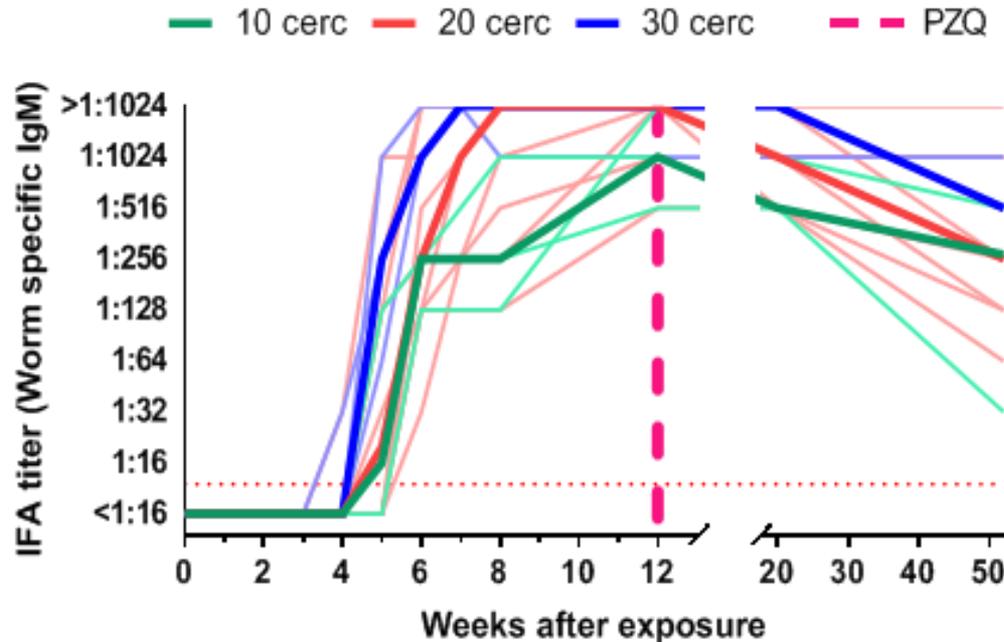
Controlled Human Schistosoma infection (CoHSI) Diagnostic aspects – antibody detection



Meta Roestenberg
and team



Healthy Dutch volunteers infected
with 10 (n=3), 20 (n=11) or 30 (n=3)
MALE *S. mansoni* cercariae



Different performance of different Ab-tests



PhD Pytsje Hoekstra

Early diagnosis and follow-up of acute schistosomiasis
in a cluster (n=34) of infected (*S. haematobium* complex)
Belgian travellers (exposed in South Africa)

PZQ



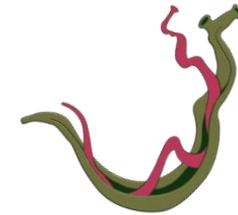
	Pre-treatment		Post-treatment
	Week 4-5 post-exposure n (%)	Week 7-8 post-exposure n (%)	Week 13-14 post-exposure n (%)
<u>Serology (Ab)</u>			
2 commercial test (ELISA and/or IHA positive) by ITM	3/33 (6%)	12/34 (35%)	11/34 (32%)
AWA-IFA (in-house LUMC)	13/33 (39%)	23/34 (68%)	25/34 (74%)



Circulating antigens

Born out of need for an alternative to microscopy

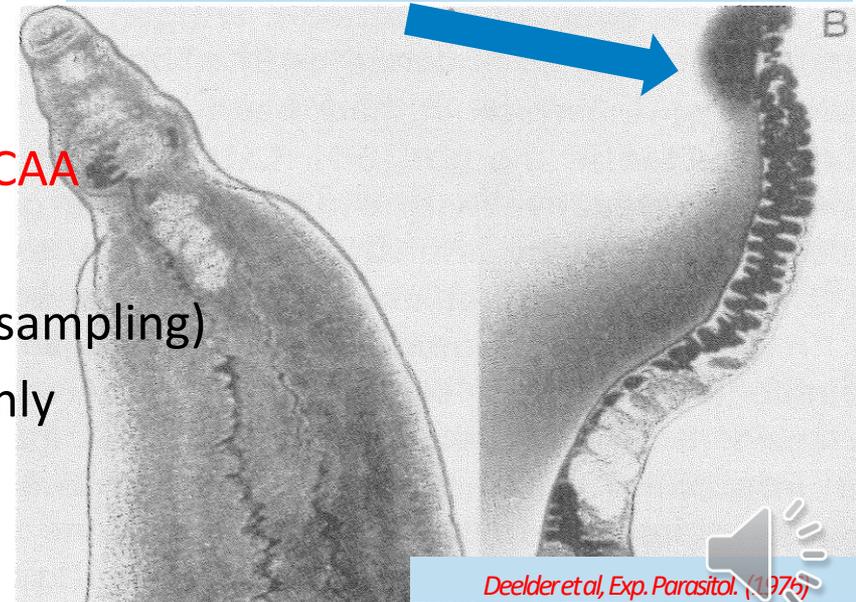
1. Detecting active infection (worm loads)
2. Highly field applicable and user friendly
3. Highly sensitive; specific
4. Case detection and for monitoring intervention



a.o.: Circulating Cathodic Antigen (CCA)
Circulating Anodic Antigen (CAA)

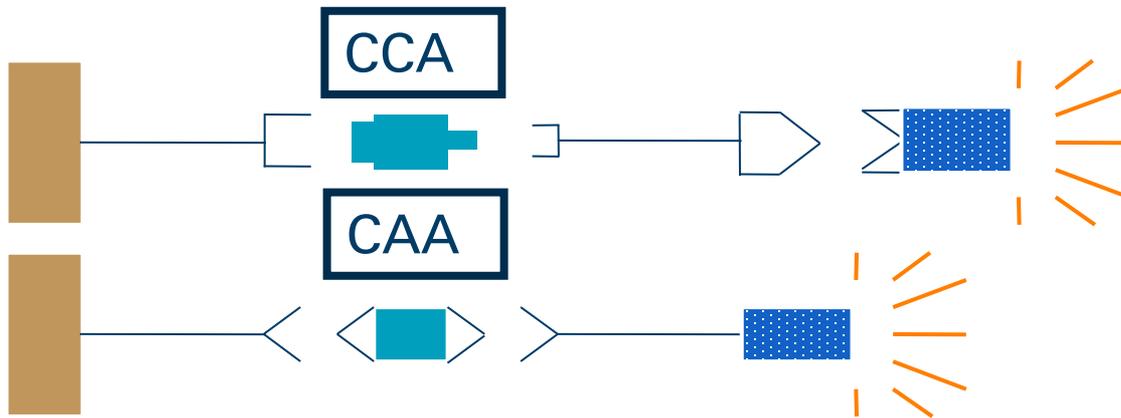
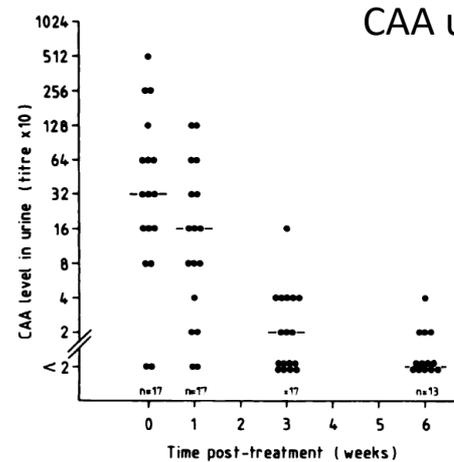
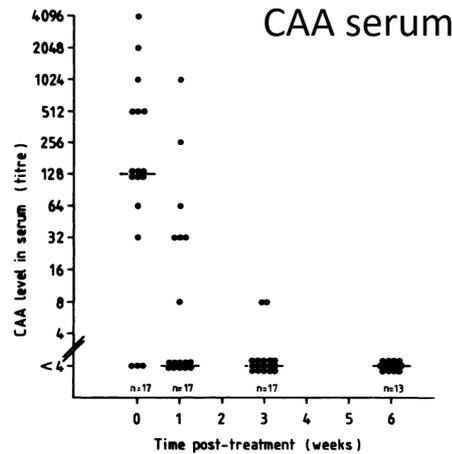
Circulating *Schistosoma* antigens CCA and CAA

- Adult worm derived glycans
- Serum => urine excreted (non-invasive sampling)
- Reflect parasite load, active infection only
- Cleared after PZQ



Deelder et al, Exp. Parasitol. (1976)

Sandwich ELISA for detection of circulating antigens



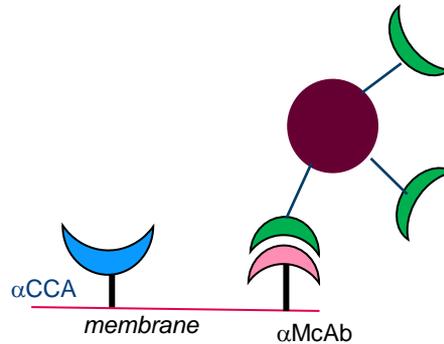
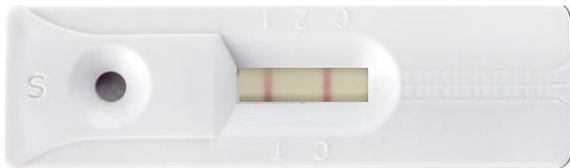
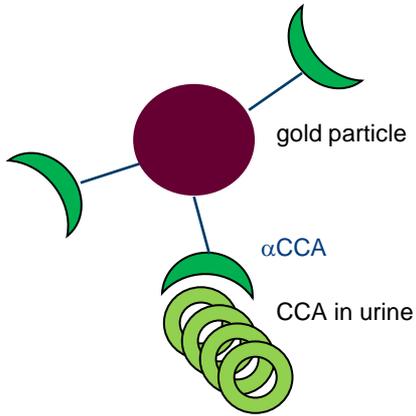
McAb - (TCA-treated) sample - McAb - enzyme substrate

Urine POC-CCA strip assay - test principle

Aim = field applicable & user friendly



Govert van Dam and team



<http://rapid-diagnostics.com/>



Silveira et al PLOS NTD 2016



Time limitation:
Exact reading at 20 min.



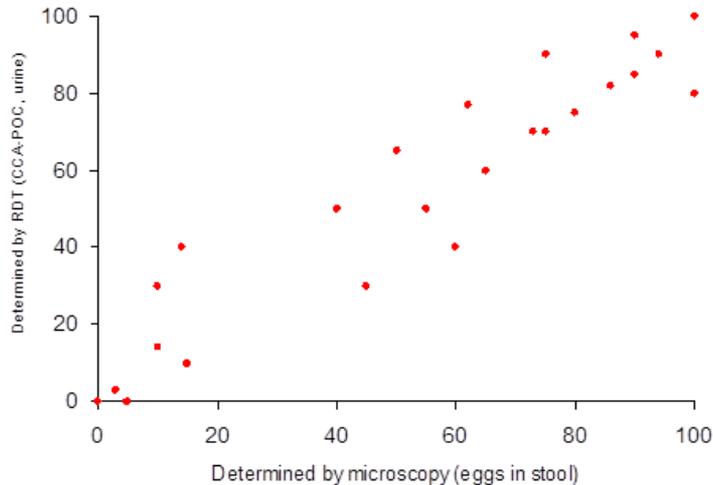
Observer dependent scoring

Urine POC-CCA strip assay => population studies

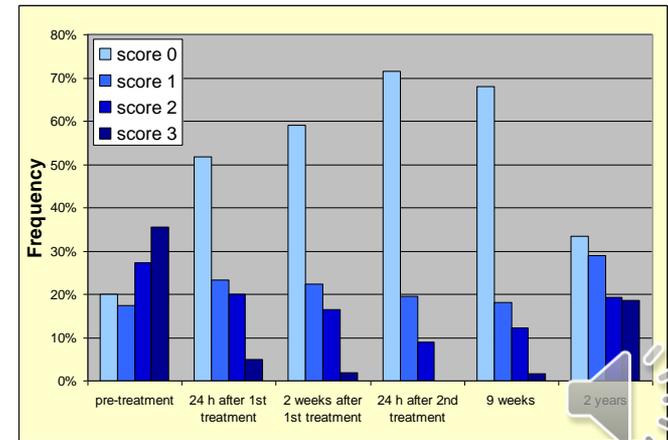
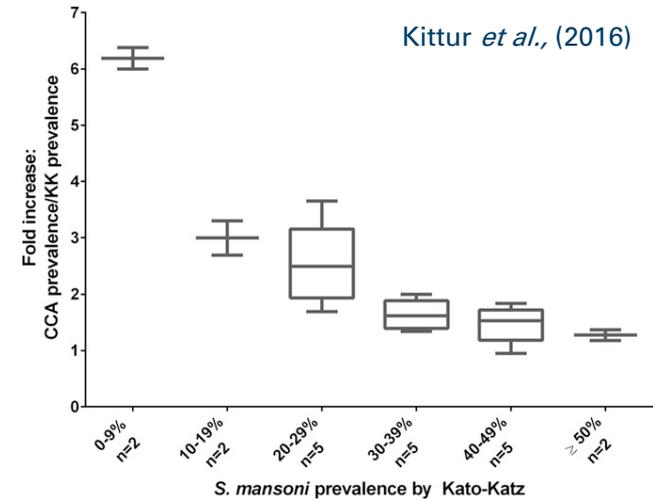
25 schools, 20-30 children per school

POC-CCA urine test vs stool microscopy

School prevalence (%)



S. mansoni region Uganda - Stothard et al., (2006)
Acta Tropica 97:219



Urine POC-CCA strip assay Recommended by WHO (2020) for *S. mansoni*



Surveys: replacement of stool microscopy by POC-CCA

Urine test; sensitivity > 1x stool microscopy
S. mansoni ; variable sensitivity for other species

Endemic settings: surveys, mapping, monitoring after treatment

Spec \approx 98% (batch variation, QC needs)

- Specificity challenges with <9M infants and pregnant women
- **Standardised reading procedures essential (reference G-scores)**



Urine POC-CCA strip assay

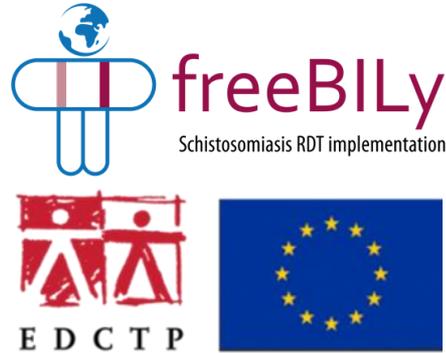
Test & treat studies, individual case detection



Govert
van Dam
and team

Implementation research in Madagascar and Gabon

www.freebily.eu



Non-endemic diagnostics????

Several studies, conflicting findings

=> Migrants with chronic *S. mansoni*

LUMC:

- CoHSI-model, not sensitive enough



Ultra-sensitive UCP Lateral Flow CAA assay

Up-Converting reporter Particle technology

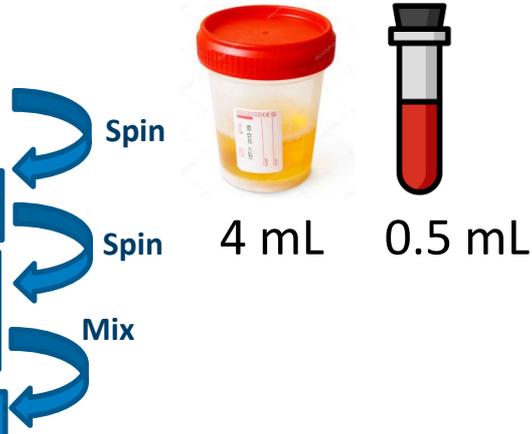
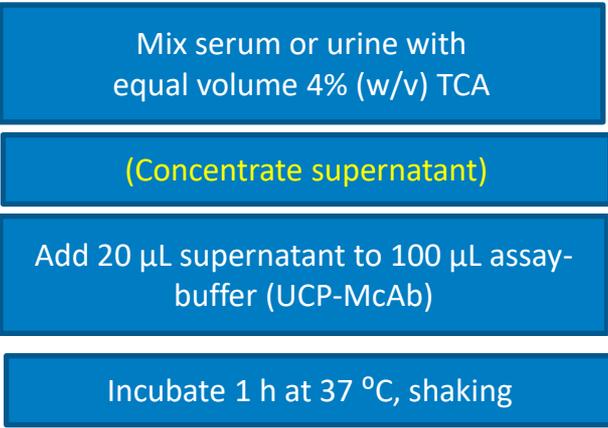


Govert van Dam and team



Paul Corstjens and team

Sample pre-treatment



Optional: dry reagents



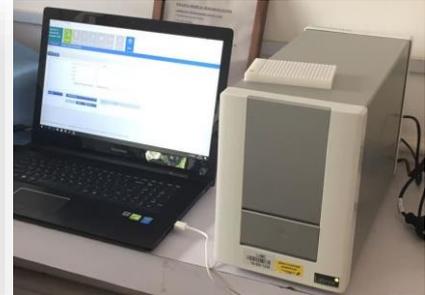
Incubate



Add LF strips, run for 1 h



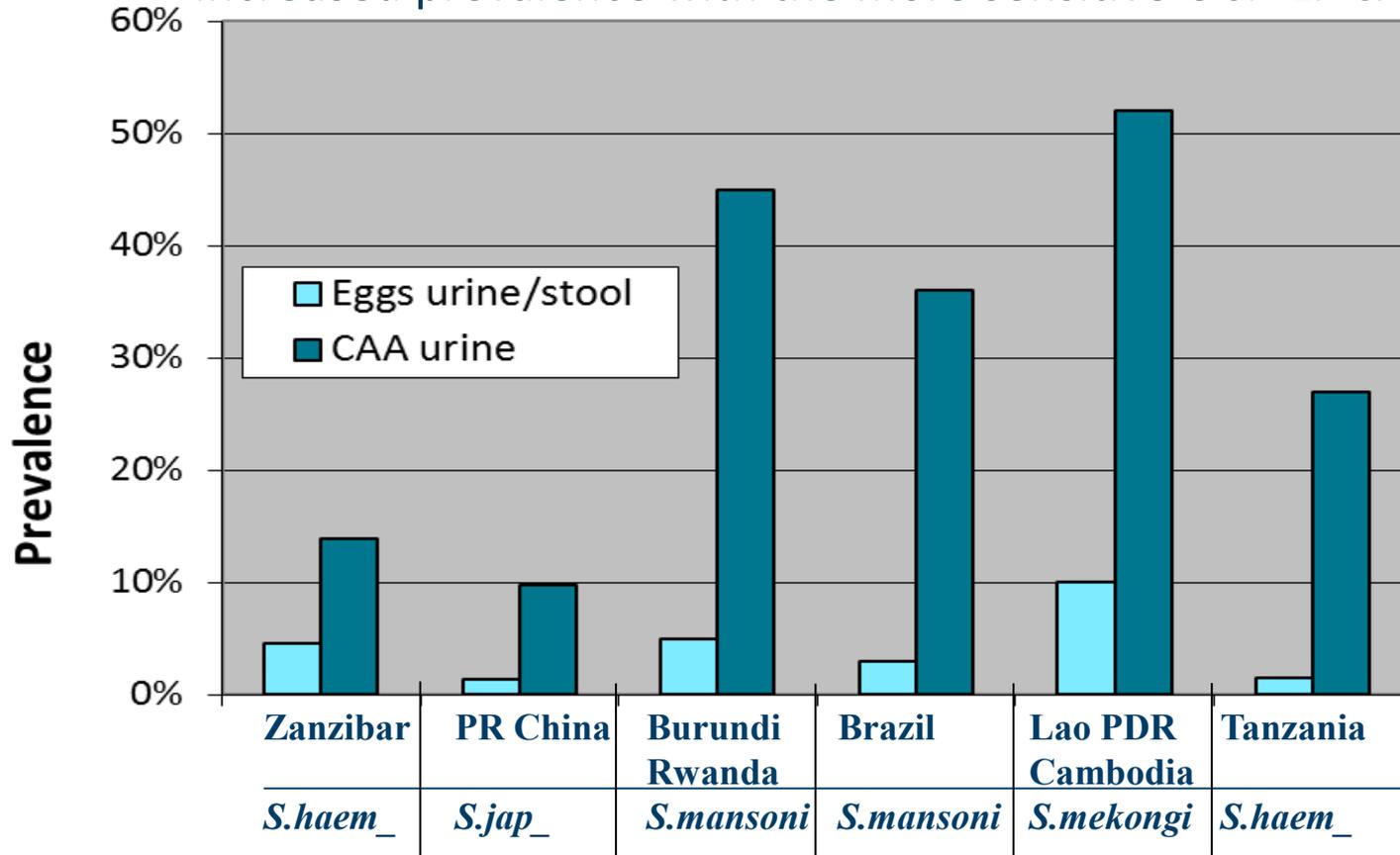
Read strips, quantitate AWA-TCA dilutions as reference standard



Real Prevalence of Infection ??

Low endemic settings by egg count:

Increased prevalence with the more sensitive UCP-LF CAA



Repeated doses of Praziquantel in Schistosomiasis Treatment (RePST) in children from Côte d'Ivoire



Govert van Dam and team

www.repst.org

Cure Rate (CR)



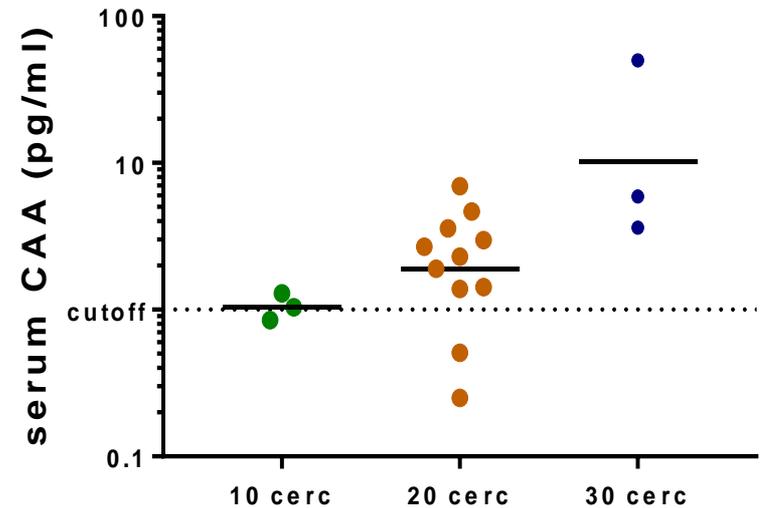
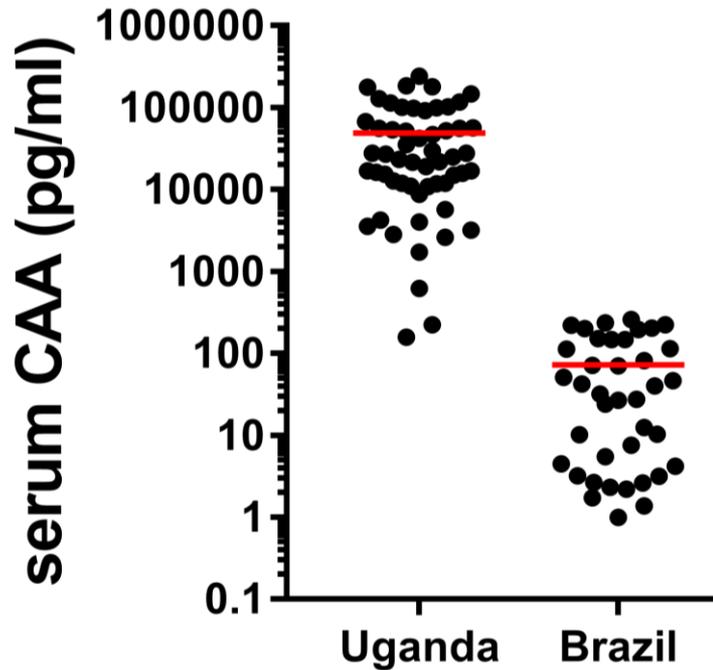
	Kato-Katz	POC-CCA	CAA (urine)	PCR (stool)
Standard (n=70)	68% (95% CI 57.1-77.6)	31% (95% CI 23.4-40.2)	23% (95% CI 16.6-31.1)	48% (95% CI 36.1-59.6)
Intense (N=83)	86% (95% CI 75.4-92.4)	36% (95% CI 26.4-46.1)	20% (95% CI 14.3-28.3)	78% (95% CI 67.6-86.0)

Intensity Reduction Rate (IRR)

	Kato-Katz			POC-CCA			CAA (urine)			PCR (stool)		
	Pre	Post	IRR	Pre	Post	IRR	Pre	Post	IRR	Pre	Post	IRR
Standard (n=70)	298 (epg)	46 (epg)	83%	9172 (pg/ml)	4265 (pg/ml)	54%	352 (pg/ml)	59 (pg/ml)	89%	61081 (Arbitrary units)	10249 (Arbitrary units)	83%
Intense (N=83)	243 (epg)	3 (epg)	95%	8488 (pg/ml)	3180 (pg/ml)	63%	338 (pg/ml)	26 (pg/ml)	93%	37085 (Arbitrary units)	317 (Arbitrary units)	99%

CAA levels in serum – different populations

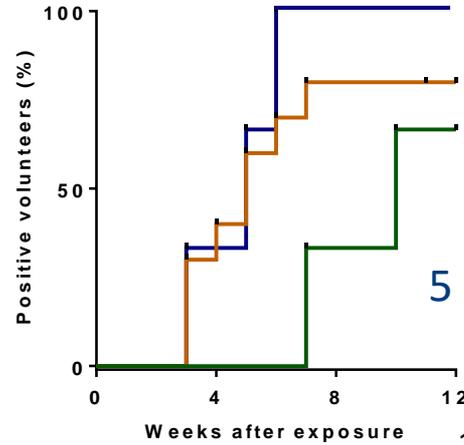
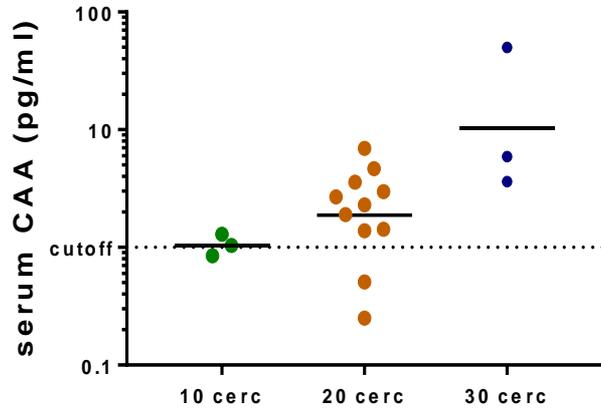
Endemic regions vs Controlled Human Infection Model



CoHSI – model

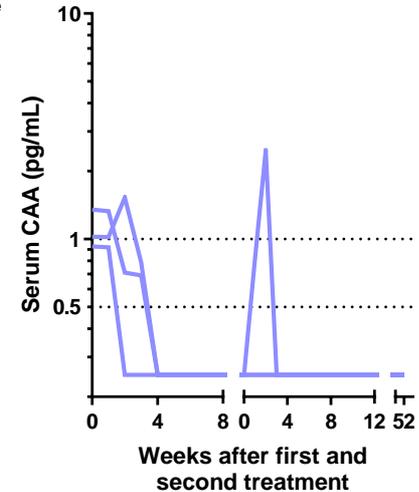
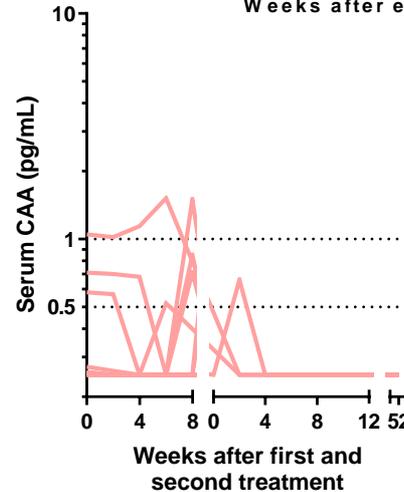
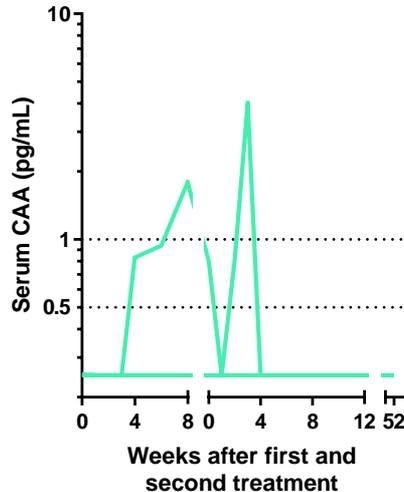
CAA in serum to monitor active infection

80% infection rate



5 weeks prepatent period

1. all: PZQ 40 mg/kgbw (2x20)
2. In 6 CAA levels persisted 3-6 weeks AT (43%): 60 mg/kgbw



Circulating Anodic Antigen detection reflecting active infection

Antigen-based diagnosis of *Schistosoma* infection: a prospective study in 106 Dutch travellers

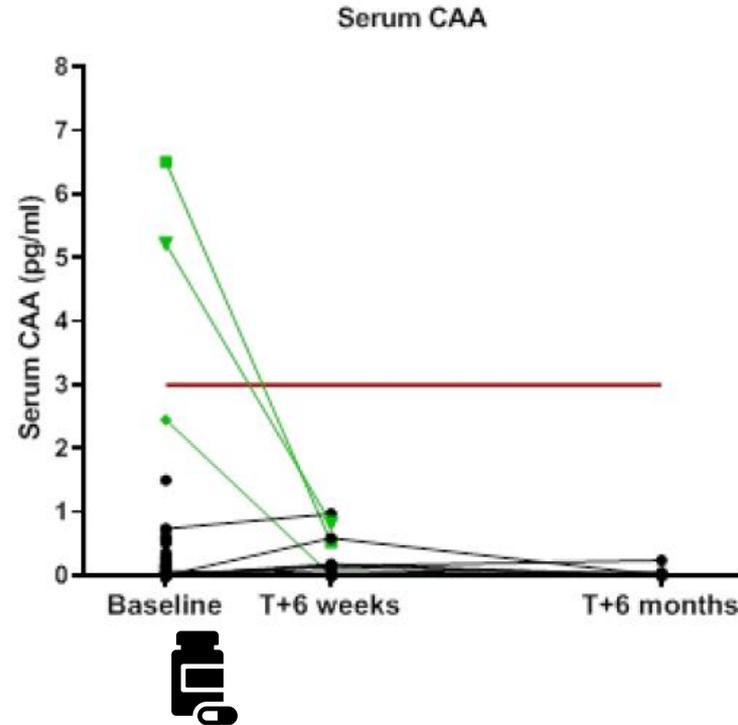
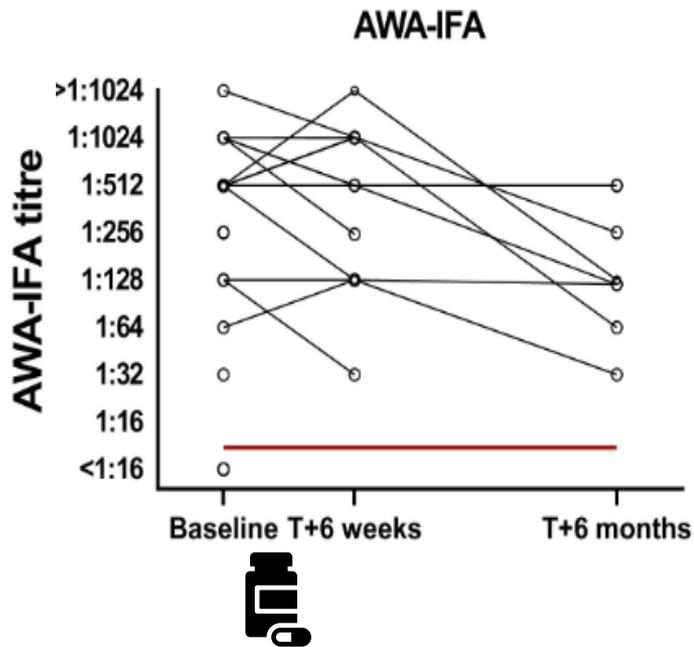


PhD Miriam Casacuberta-Partal

22 Ab positive

–

2 serum CAA (dry assay) positive

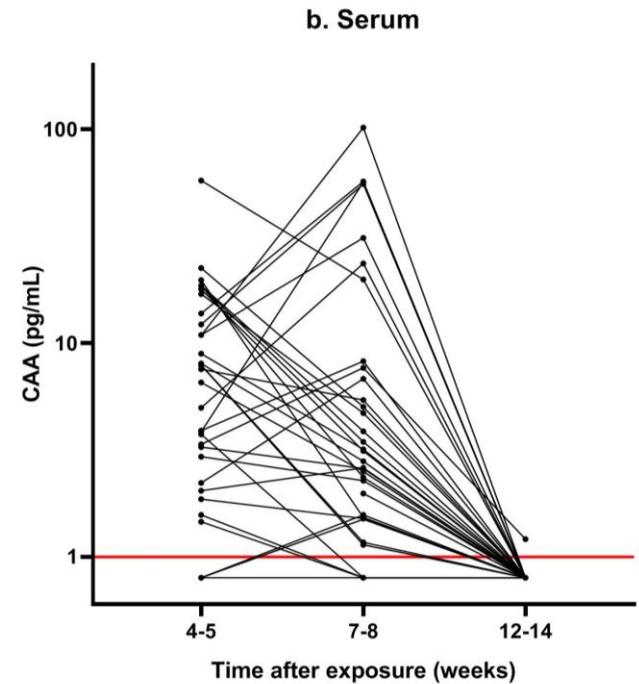
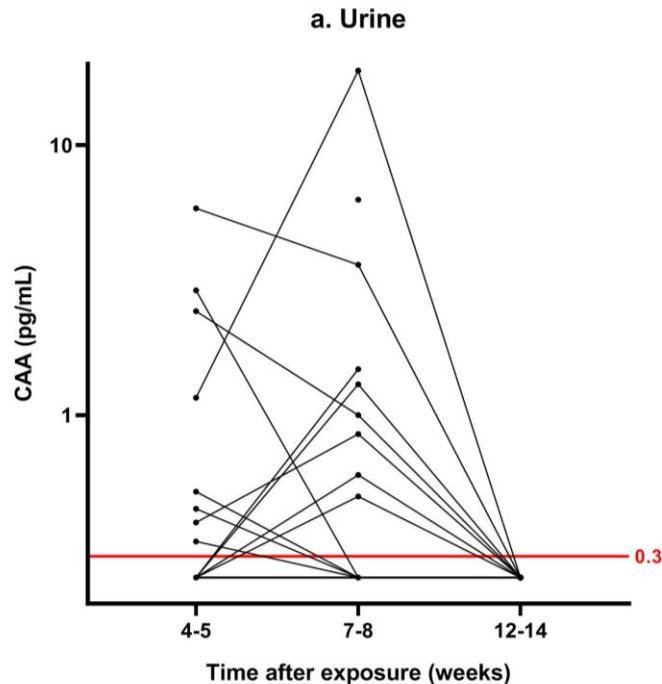


Circulating Anodic Antigen detection reflecting active infection



PhD Pytsje Hoekstra

Early diagnosis and follow-up of acute schistosomiasis in a cluster (n=34) of infected (*S. haematobium* complex) Belgian travellers (exposed in South Africa)



Summary: from discovery to implementation

No “one size fits all” test

DNA detecting tests (NAAT)

- Increasingly replaces microscopy; FGS/MGS diagnosis

Future: field-friendly test formats? LAMP, RPA etc.

Serology (Ab testing)

- With the right test: early detection in acute schistosomiasis

Circulating antigens

- POC-CCA: S.m. chronic infections (QC in place)
- Ultra-sensitive UCP-LF CAA: early detection, post-PZQ
- UCP-LF CAA for trials/research; pooled samples
- **Limited availability**

- <https://youtu.be/juqlvxN6u0Y>



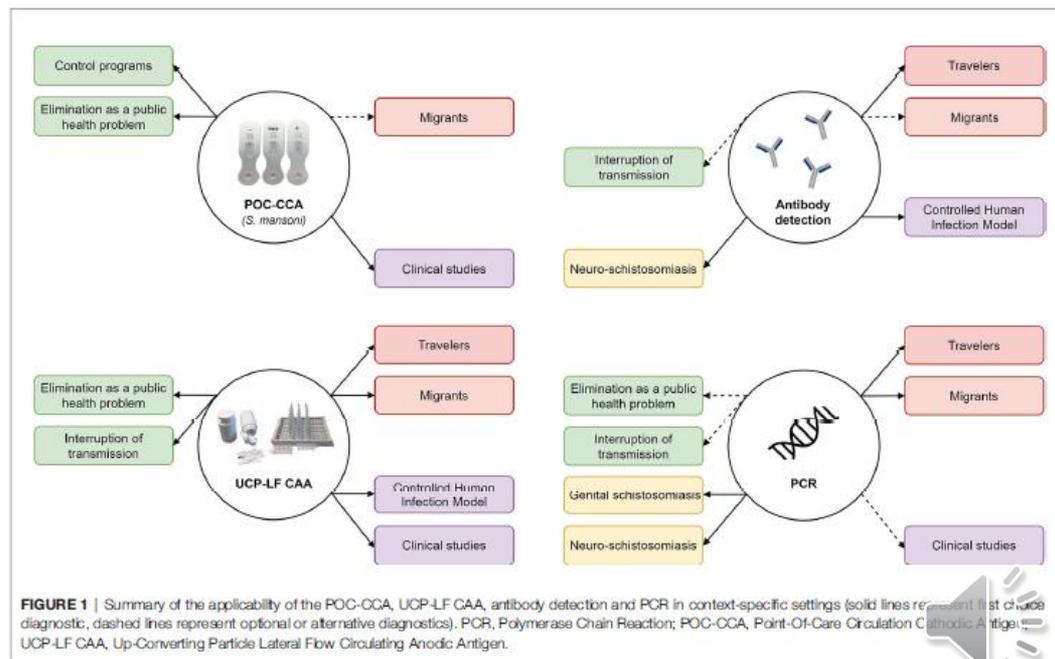
Context-Specific Procedures for the Diagnosis of Human Schistosomiasis – A Mini Review



Pytsje T. Hoekstra*, Govert J. van Dam and Lisette van Lieshout

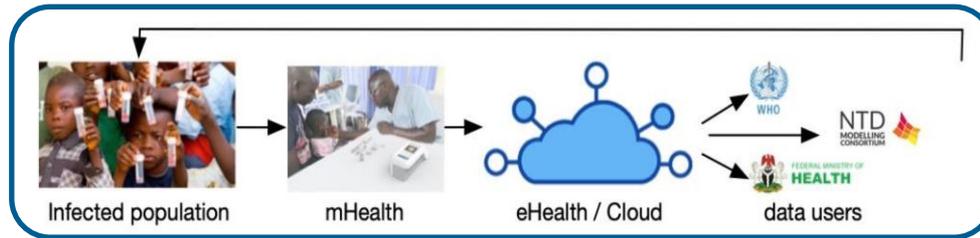
Department of Parasitology, Leiden University Medical Center, Leiden, Netherlands

OPEN ACCESS



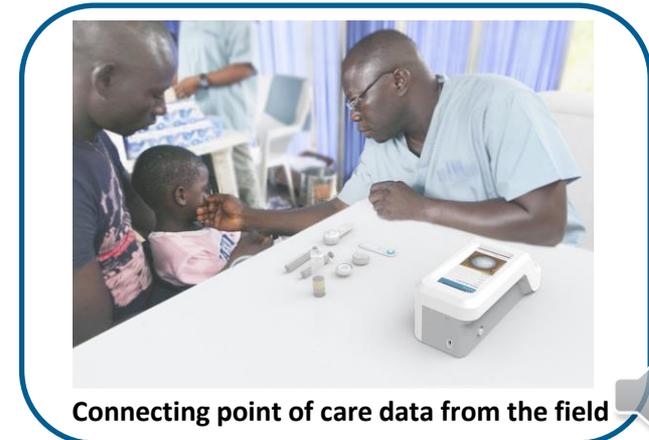
Future steps in CCA/CAA diagnostic research

- Basic biology (structure, function)
- Kinetics (host, infection, post-treatment)
- Operational (pooling; data connectivity)



RE – ASSURED – diagnostic requirements

- R**Real-time connectivity
- E**ase of specimen collection
- A**ffordable
- S**ensitive
- S**pecific
- U**ser-friendly
- R**apid and Robust
- E**quipment-free
- D**eliverable





Leiden University
Medical Center



Part of the LUMC Schisto / CoHSI team

Thank you!

CoHSI - volunteers



freeBLy
Schistosomiasis RDT implementation

Prof. Dr. Flu foundation

EDCTP

